

Testing Methodologies Applicable to Shiga Toxin-Producing Escherichia Coli (STEC)

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Shiga Toxin-Producing Escherichia Coli (STEC)



Enterohemorrhagic E. coli (EHEC)

- Gram-negative rods
- 400+ STEC serotypes (not all pathogenic)

How to define when a given strain is or is not pathogenic?

It is very difficult to distinguish STEC from other E. coli as the only phenotypic feature that identifies STEC is the production of the Shiga toxins





Strains with co-occurance of 2 virulence factors: Toxin (stx1/2) and Adhesion determinants (eae)



EHEC (Highly Pathogenic STEC)



Potentially pathogenic bacteria *stx* + and *eae* +

Leading cause of EHEC infections in humans -> O157:H7 serotype

Most common non-O157 serogroups

- → O26, O103, O91, O146 and O145 (Europe)*
- → O26, O45, O103, O111, O121 and O145 (US)*



Food Matrices at Higher Risks?

Raw or undercooked meat, Raw milk products, Fruit & Vegetables (e.g. sprouts)

 Table 4:
 Number of human cases, hospitalisations and deaths per implicated food vehicle category reported in strong evidence STEC food-borne outbreaks from 2012 to 2017

Implicated food vehicle category (number of reported strong evidence outbreaks; number of reporting countries)	Human cases	Hospitalisations	Deaths
Bovine meat and products thereof (15; 7)	143	76	0
Milk and dairy products ^(a) (14; 8)	94	43	2
Tap water, including well water (8; 4)	75	7	0
Vegetables, fruit and products thereof ^(b) (7; 3)	575	73	2
Pig meat and products thereof (2; 1)	6	2	0
Other or mixed red meat and products thereof (2; 2)	10	0	0
Sheep meat and products thereof (1; 1)	27	9	0
Unspecified meat ^(c) (1; 1)	2	1	0
Fish and seafood ^(d) (1; 1)	5	0	0
Herbs and spices (1; 1)	50	3	0
Total	987	214	4

(a): Includes all foods categorised under 'milk', 'dairy products (other than cheeses)' and 'cheese' from the Zoonoses Catalogue. In at least six outbreaks, the actual source was raw milk.

(b): Includes all foods categorised under 'fruit, berries and juices and other products thereof' and 'vegetables and juices and other products thereof' under the Zoonoses Catalogue.

(c): All foods categorised under 'meat and meat products' from the Zoonoses Catalogue.

(d): Includes all foods categorised under 'fish and fish products' and 'crustaceans, shellfish, molluscs and products thereof'.





STEC genome plasticity → Emerging strains

➔ The detection of the stx gene(s) is the only true discriminant between STEC and other E. coli.

Current testing methods \rightarrow High percentage of presumptive positive samples that could take up to 5 days for culture confirmation

- Immunological methods, based on the detection of the Shiga toxins, provide indirect evidence of the presence of STEC
- Methods based on **PCR** are the **most appropriate** approaches to detect STEC in complex matrices.

 \rightarrow It is possible to identify accessory virulence features, such as the adhesion determinants (eae gene),

 \rightarrow It gives an indication of the presence of STEC strains considered to be more likely to cause severe disease in humans.



Pathogenic E.coli Confirmation Test Volume Rapid Methods

Globally, 41M tests are performed / 15M of these tests are performed using rapid methods (PCR, immunoassay, lateral flow, etc.)

→ Approximately 6-8M tests for confirmation after rapid method screening

Pathogenic E.coli Test Volume – Rapid Methods

Pathogen *E.coli* Confirmation Test Volume – Rapid Methods





Bio-Rad solution for STEC detection in food





iQ-Check STEC solution by Bio-Rad



iQ-Check STEC VirX / iQ-Check STEC SerO



Description:

- Based on gene amplification and detection by real-time PCR
- Ready-to-use PCR reagents contain oligonucleotides (primers and probes) specific for *stx1/stx2* and *eae* virulence genes, as well as DNA polymerase and nucleotides
- iQ-Check STEC SerO for TOP 7 serogroups
- Included a synthetic DNA internal control and positive and negative control
- Detection and data analysis are optimized for use with CFXOpus DW & CFX Manager IDE





iQ-Check PCR Real Time STEC kit - Workflow



Enrichment:

 Different enrichment proposed for differents matrices





Extraction and plate preparation step:

- Flexible solution: from low to high throughout
- One plate & one protocole for differents pathogenes
- Easy to use and implement in laboratories

Amplification step:

- System easy to install
- User friendly software: only on window to set up and run PCR
- All iQ-Check food test can be run on the same plate



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International Validation

- AOAC, Microval, approved
- Screening step for the USDA MLG reference method Chapter 5C.02.

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dd-PCR STEC solution by Bio-Rad







2nd generation

PCR Qualitative

Real-time PCR Relative Quantification





Droplet Digital PCR (ddPCR) – Power is Partitioning



- Partition sample so **individual nucleic acids** are localized to separated containers (nanoliter droplets)
- PCR reactions are independent, single amplification events
- The number of negative/positive droplets is **directly related to the initial concentration** in the sample
- Co-localization of genes (ex. For STEC stx and eae)

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ddPCR – Propositions



Bacterial isolation allowing colocalisation caracterisation



BIO RA





- Flexible solution: from low to high throughout
- Ready-to-use PCR reagents
- Specific for *stx1/stx2* and *eae* virulence genes and TOP 7 serogroup
- Solution adapted for differents matrices
- Easy to use and implement in laboratories
- User friendly software and automated interpretation
- One plate & one protocole for differents pathogenes
- International Validation: AOAC, microval and USDA FSIS











Thank you for your attention!!!

